Influence of Mussel Aquaculture on Nitrogen Dynamics in a Nutrient Enriched Coastal Embayment

Peter J. Cranford^{1*}, Peter M. Strain¹, Michael Dowd², Barry T. Hargrave¹, Jonathan Grant³, Marie-Claude Archambault³

Running Head: Mussel aquaculture and coastal nitrogen dynamics

 ¹ Ecosystem Research Division, Fisheries and Oceans Canada, Bedford Institute of Oceanography, PO Box 1006, Dartmouth, Nova Scotia, Canada B2Y 4A2
 ² Department of Mathematics and Statistics, Dalhousie University, Halifax, Nova Scotia, Canada B3H 3J5

³ Department of Oceanography, Dalhousie University, Halifax, Nova Scotia, Canada B3H 4J1

Emails: *cranfordp@mar.dfo-mpo.gc.ca

1 ABSTRACT: The combined influences of intensive mussel aquaculture and watershed 2 nutrient inputs on nitrogen dynamics in Tracadie Bay (Prince Edward Island, Canada) 3 were examined using a nitrogen budget and an ecosystem model. Budget calculations 4 and inputs and parameters for the model were based on extensive field data. Both 5 approaches showed that mussel aquaculture has a dominant influence on all aspects of the 6 nitrogen cycle and dramatically alters pathways by which nitrogen reaches the 7 phytoplankton and benthos. A large proportion of phytoplankton production is supported 8 by land-derived nitrogen and this anthropogenic input is important for sustaining existing 9 levels of mussel production. The amount of nitrogen removed in the mussel harvest is 10 small compared to agricultural nitrogen inputs and the amounts excreted and 11 biodeposited on the seabed. Mussel biodeposition greatly increases the flux of nitrogen to 12 the benthos, with potentially serious eutrophication impacts. Results from the observation 13 based nitrogen budget and dynamic model were compared and both support the above conclusions. However, the ability of the model to test different scenarios and to provide 14 15 additional information (e.g. fluxes) over a finer spatial scale led to insights unattainable 16 with a nitrogen budget. For example, food appears to be less available to mussels at the 17 head of the Bay than at the mouth, despite the lower density of grow-out sites in the 18 former location. The number of fundamental ecosystem processes influenced by the 19 mussels and the complexity of their interactions makes it difficult to predict the effects of 20 mussels on many ecosystem properties without resorting to a model.

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22 KEY WORDS: eutrophication, nitrogen cycling, nitrogen budget, ecosystem model,

23 ecophysiology, biodeposition, excretion

24 INTRODUCTION

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26 Natural and farmed populations of suspension-feeding bivalve molluscs exert a 27 dominant influence on energy flow and nutrient cycling in many coastal marine 28 ecosystems, particularly in inlets where water residence time is long and bivalve biomass 29 is high (Smaal and Prins 1993; Dame 1996; Dame and Prins 1998; Cranford et al. 2003; 30 Newell 2004; Grant et al. 2005). By creating structurally complex shell habitat and 31 performing a wide array of ecological functions, bivalve populations can substantially 32 modify benthic and pelagic communities at different trophic levels and alter energy flow 33 and nutrient cycling over the scale of entire coastal ecosystems. Potential mechanisms for 34 ecosystem effects include the utilization of particulate food resources by the bivalves, the 35 biodeposition of faeces and pseudofaeces, and the excretion of metabolites. Bivalve 36 aquaculture is expanding rapidly in many countries and a comprehensive understanding 37 of the influence of this industry on coastal ecosystems, as well as interactions with other 38 anthropogenic stressors, is fundamental for developing strategies for the sustainable 39 management of the coastal zone as well as the aquaculture industry.

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41 Dense bivalve populations and communities are known to influence the nitrogen 42 cycle in coastal ecosystems with the degree of control depending largely on site-specific 43 hydrographic conditions (Dame 1996; Newell 2004). Bivalves exert "bottom-up" nutrient 44 control on the phytoplankton by (1) the excretion of large amounts of nitrogen (primarily 45 ammonia) and (2) by depositing organic matter from ingested phytoplankton and detritus 46 (also includes remnants from ingested auto- and heterotrophic microplankton and 47 zooplankton), which facilitates the benthic recycling of nitrogen. The increased organic 48 loading of sediments from biodeposition may enhance the retention of nutrients, coming

49 from both the sea and land, in coastal systems and stimulate mineralization and nitrogen 50 release rates (Newell 2004; Nizzoli et al. 2006). Nitrogen fluxes from the recycling of 51 biodeposits trapped within suspended bivalve culture ropes and other structures are also 52 ecologically significant and can be higher than benthic fluxes (Mazouni 2004; Richard et 53 al. 2006: Nizzoli et al. 2006). Accelerated nitrogen cycling and coastal nitrogen retention 54 directly attributed to bivalve excretion and biodeposition may significantly accelerate 55 phytoplankton turnover and production (Doering and Oviatt 1986; Doering et al. 1989; 56 Asmus and Asmus 1991; Prins et al. 1995).

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58 Coastal ecosystems are increasingly stressed by many human activities and the 59 potential effects of aquaculture should not be considered in isolation. Significant 60 ecosystem-level interactions are expected between bivalve aquaculture and 61 eutrophication (Dame 1996; Cloern 2001; Newell 2004). Applications of agricultural 62 fertilizer to farm lands enrich nutrient concentrations in surface and ground water. Upon 63 reaching coastal systems, these nutrients stimulate plant growth and can disrupt the 64 natural balance between the production and metabolism of organic matter. Large 65 populations of bivalve filter-feeders are believed to control coastal ecosystem responses 66 to nutrient loading by ingesting large quantities of microalgae and thereby increasing the estuary's grazing, or "top down", control of excess phytoplankton biomass (Dame 1996; 67 68 Cloern 2001; Newell 2004). In addition to potentially having the capacity to clear excess 69 phytoplankton from suspension, it has been suggested that bivalve aquaculture may help 70 ameliorate the impacts of nitrogen enrichment in eutrophic coastal waters by removing 71 excess nitrogen in the shellfish harvest (e.g. Rice 2000; 2001). This has led to suggestions 72 that shellfish aquaculture be incorporated in a nutrient trading system as an alternative to 73 nitrogen reduction for improving coastal water quality (Lindahl et al. 2005).

75 The number of ecosystem processes potentially influenced by bivalve culture and the 76 complexity of their interactions (e.g. simultaneous top down and bottom up controls on 77 phytoplankton) makes it difficult to predict the effects of the bivalves on many ecosystem 78 properties. Such predictions are further complicated by ecological interactions between 79 bivalve suspension feeders and eutrophication (Cloern, 2001). The present study was 80 conducted to further scientific understanding of the nitrogen dynamics of a coastal 81 aquaculture embayment receiving nutrient inputs from land-use. Two different 82 approaches were applied to analyzing major elements of the nitrogen cycle; an 83 observation based nitrogen budget and a dynamic ecosystem model. Results were 84 compared to provide insights into the individual strengths and limitations of each 85 approach with respect to their possible applications. A related objective was to apply 86 these approaches to test hypotheses and refine theories including; (1) the potentially 87 dominant role of mussel aquaculture in nitrogen dynamics at the coastal ecosystem scale, 88 (2) the influence of mussel culture on coastal nutrient retention, and (3) the capacity of 89 the mussel harvest to ameliorate impacts from nitrogen enrichment.

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91 The site used in this study is Tracadie Bay (Fig. 1), one of the more extensively 92 leased mussel aquaculture inlets in Prince Edward Island (PEI; Fig. 1). PEI coastal inlets 93 supply 77% of the \$30 million total value of the mussel culture industry in Canada (DFO 94 2005). Many PEI embayments, including Tracadie Bay, receive agriculture run-off, and 95 nutrient inputs from fertilizers have resulted in eutrophic conditions (Raymond et al. 96 2002). Tracadie Bay has been the focus of several studies and extensive field sampling 97 programs have been conducted to document the physical oceanography (Dowd et al. 98 2001; 2002) and biogeochemistry of the Bay and adjacent waters (e.g. Bates and Strain

99	2006). The bio-physical control of the distribution of phytoplankton in Tracadie Bay has
100	been examined using a simple tracer model (Dowd 2003), a lower trophic level
101	ecosystem box model (Dowd 2005) and a spatially explicit ecosystem model of seston
102	depletion by the mussel culture (Grant et al. submitted). All three approaches suggest that
103	mussel grazing strongly affects phytoplankton levels in this inlet, and that the spatial
104	pattern is also dictated by water motion and nitrogen run-off. Dowd (2005) developed an
105	ecosystem box model approach and conducted a preliminary examination of how mussels
106	affect nitrogen cycling in the bay. This approach was further refined for the present study
107	and applied to quantitatively describe nitrogen dynamics dictated by the major interacting
108	ecosystem components (nutrients, phytoplankton, mussels, detritus and benthos).
109	Detailed biological and chemical field data for Tracadie Bay, which were not available
110	during initial model development, are compared to model output. Whenever possible,
111	actual field data are used in the budget calculations and to determine parameters and
112	initial and boundary conditions for modelling purposes.
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Information on the Tracadie Bay drainage basin and land-use was supplied by the PEI
Department of Environment for calculation of drainage basin and land type areas using
GIS (ArcInfo Ver. 9.1). The Tracadie Bay watershed (Fig. 2) totals 146.2 km² and land
types include forest (46.8% of total area), agriculture (32.7%), wetlands and beach
(12.5%), and urban and other (8.0%). The Winter River watershed (Fig. 2) is the largest
drainage sub-basin to the Bay (69.7 km²) and a relatively large fraction (41.4%) is used
for agriculture (grain, potato, hay and pasture).

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133 Water Chemistry. Water samples were collected by Niskin bottle from 1 and/or 3 m 134 depth at 12 stations in Tracadie Bay (circles in Fig. 2) approximately once per month in 135 the ice-free seasons (June to November) for two years (2002-2003). Some limited sampling was also conducted through the ice in winter months. Additional nutrient data 136 were available from samples collected in a previous program conducted in 1998-99. 137 138 which included sampling in the months July through October. Vertical profiles with a 139 portable CTD (SeaBird 25) provided supporting salinity and temperature data. Dissolved 140 inorganic nutrients were determined for all samples, using standard autoanalyzer 141 techniques for nitrate and nitrite (Strain and Clement 1996), and ammonia (Kérouel and 142 Aminot 1997). In this paper we will use 'nitrate' to refer to the total oxidized inorganic 143 nitrogen (nitrate + nitrite), and TIN (total inorganic nitrogen) to refer to nitrate + nitrite + 144 ammonia. Nutrient samples were also collected from eight stations in the Winter River 145 (squares in Fig. 2) approximately weekly from May to November 2003 by hand dipping 146 sample bottles into the surface layer from shore.

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148 Suspended particulate matter (SPM) in the water samples was collected on 1.7 um 149 nominal pore size glass fibre filters (25 mm diameter Micro Filtration Systems type GC). 150 SPM was collected in triplicate on prewashed, precombusted (450 °C for 4 h), tared glass 151 fibre filters. SPM levels were determined after rinsing the filters under vacuum with 152 isotonic ammonium formate to remove salt and drying the filters at 60°C and weighing to 153 the nearest 0.01 mg. Particulate organic matter (POM) concentration was determined as 154 total weight loss upon ignition at 520°C for 6 h and the organic fraction (f_{POM}) was 155 calculated as POM/SPM. Samples to characterize the SPM and organic matter were 156 provided from the Niskin water sampling surveys described above and using a moored 157 Water Transfer System (McLane Research Laboratories, Inc., Falmouth MA, USA) that 158 filters water in situ at programmed intervals onto glass fibre filters (47 mm diameter 159 Micro Filtration Systems type GC). This system made it possible to collect frequent SPM 160 samples for the evaluation of f_{POM} . Chlorophyll *a* in SPM samples collected on glass fibre 161 filters (same type as above) during the Niskin survey was determined from the in vitro 162 fluorescence (Turner Designs fluorometer calibrated against pigment from spinach) of 163 90% acetone extracts of the filtered material.

164

Nitrogen Cycle. Table 1 lists the important reservoirs, internal fluxes, and external inputs and outputs that are elements of the nitrogen cycle in Tracadie Bay. The letters identifying the reservoirs in Table 1 will be used throughout this paper. Note that the TIN reservoir is distinguished from other reservoirs and fluxes expressed in nitrogen equivalents. Table 1 also shows which components are quantified in the nitrogen budget and modelling approaches.

171

Whenever possible, we have used field data for constructing the nitrogen budget and for setting boundary / initial conditions for the models and assessing their performance. For many such purposes, we synthesized the available field data and produced seasonal cycles (monthly) using objective analysis. Objective analysis is sometimes referred to as an 'optimal estimator', because the Gauss-Markov theorem on which it is based claims that "Given the statistics of the field being measured and the noise levels involved, no other [linear] analysis could perform better" (Bretherton et al. 1976).

179

180 To characterize the nitrogen cycle, we require estimates of the nitrogen levels in the 181 phytoplankton (P), dissolved nutrients (TIN) and detritus (D) reservoirs within Tracadie 182 Bay and for the offshore. We have formulated both the nitrogen budget and the models 183 to use a common currency for the different ecosystem reservoirs and fluxes, expressing 184 all quantities in nitrogen equivalents. For many quantities, we use units of tonnes nitrogen or tonnes nitrogen per vear (t N or t N y⁻¹). P was determined from the 185 186 chlorophyll a distribution, converted to nitrogen using a carbon:chlorophyll a ratio of 50, 187 and a Redfield C:N ratio of 106:16 (molar) in the phytoplankton. The TIN values require 188 no conversion, but require summing the nitrate and ammonia concentrations. The detritus 189 is considered here to be the fraction of organic matter not associated with living

phytoplankton cells. Since most of the living cells in the water column are
phytoplankton, the amount of nitrogen in detritus (N_D) can be calculated from the
difference between nitrogen in the total particulate organic matter (N_{POM}) and the
nitrogen in the phytoplankton (N_P):

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$$N_{\rm D} = N_{\rm POM} - N_{\rm P}.$$

N in bacteria, either free-living in the water column or associated with detritus, is notconsidered in these calculations.

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198 For areas inside Tracadie Bay, we have sufficient data for POM and P to calculate 199 N_D. P is converted to N_P as described above; N_{POM} is calculated using the Redfield C:N 200 ratio and 1.9 as a typical ratio of organic matter:organic carbon. These N_D values were 201 then objectively analyzed in the same way as the other parameters. However, the extra 202 steps in the calculation add to the uncertainty of the resulting seasonal cycle. For the 203 offshore station, sufficient POM data are not available for this calculation. Instead, we 204 have used the observation that N_P+N_D is approximately constant (~6.4 M N) in data 205 from the Tracadie Bay area to approximate offshore D from the offshore seasonal cycle 206 of P. Although the determination of the nitrogen in both P and D are operational, they 207 are internally consistent because the same filters to collect SPM were used in both 208 measurements.

P, TIN and D levels in offshore waters influencing Tracadie Bay are boundary conditions required for both the nitrogen budget and the models. Seasonal cycles for these quantities were predicted by objective analysis for the offshore region using both data from this study at Station W12 (Fig. 2) and data from the BioChem data archive maintained by the Department of Fisheries and Oceans for adjacent areas of the Magdalen Shallows in the Gulf of St. Lawrence. Concentrations were predicted for 5 m 215 depth at W12 for the middle of each month. Julian day is used for the time axis (i.e. data 216 from all years are merged), and the data set is expanded to cover the range 0 to 365 days 217 ± 182 days to avoid biasing estimates at each end of the calendar year. Fig. 3 shows one 218 such seasonal cycle predicted for nitrate at Station W12, and the distribution of data 219 points on which it is based (~ 1400 measurements). The corresponding seasonal cycles 220 for ammonia and chlorophyll were also determined (not shown), and these cycles were 221 used to calculate seasonal cycles for P, TIN and D in nitrogen equivalents as described 222 above.

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Objective analysis was also used to estimate seasonal cycles for areas within Tracadie Bay for comparison with model predictions and for calculating mean nitrogen inventories for the nitrogen budget. Estimates have been made for each box used in the model described below. For these analyses, the seasonal cycle was predicted for each point on a 200 m grid in Tracadie Bay. Data for each month for each point in each model box were then averaged to produce a seasonal cycle for the model box. For example, Fig. 4 shows the predicted chlorophyll cycle for the boxes of the lower trophic level model.

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The seasonal cycles predicted in this way for both the offshore and the model boxes within Tracadie Bay describe conditions that are averaged over all the available data and do not describe a specific year's annual cycle. The temporal and spatial distributions of these data vary between the different model boxes and the offshore. Some gaps in the sampling exist, such as during the spring due to ice break-up. This limitation is most serious with TIN, for which the few available winter measurements are highly variable (7 - 150 M) with a mean value (52 M) that is much higher than observed at other times 239 of year. Although all these estimates are based on data, they are still idealized

representations of the annual cycle.

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242 Lower Trophic Level Model. Dowd (2005) developed a lower trophic level 243 ecosystem model for Tracadie Bay, implemented with three spatial boxes (Fig. 5), that 244 predicts the levels of phytoplankton (P), zooplankton, dissolved nutrients (TIN) and 245 detritus (D), as well as their interactions with a simplified benthos (B) that includes terms for particle settling, permanent burial, resuspension and nutrient remineralization. Water 246 247 exchange coefficients were derived from a heat budget calculation using observed 248 temperature time series. The impacts of mussel aquaculture (M) are evaluated by 249 superimposing the grazing activity of the mussels onto this system. The elements of the 250 nitrogen cycle described by the model are listed in Table 1. As used in this study, the model formulation differs from that of Dowd (2005) in the following ways: 251 252 The units have been converted to nitrogen equivalents. ٠ 253 Since tests showed that the zooplankton pelagic state variable (Z) had little impact ٠ 254 on the nitrogen dynamics, it has been eliminated. Ecosystem closure is achieved by a quadratic loss term $_{\rm P}{\rm P}^2$ that represents mortality and grazing of 255 256 phytoplankton. This loss term is put back into the detritus pool, resulting in the 257 following equation for P (which replaces Eq. 1 in Dowd, 2005): $\frac{dP}{dt} = f\{N;k_n\}\gamma_P P - \lambda_P P^2 - I_m P + K(P_\infty - P)$ 258 259 The corresponding zooplankton terms have been dropped from the equations for 260 N and D. D has been adjusted for the flux from P. and the equation for B is 261 unchanged.

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262	• The convolution integral that governed benthic remineralization has been replaced
263	with a simple temperature and B dependent efflux (rather than being based on the
264	weighted time history of the input flux).
265	• The external N source has been replaced with a seasonally variable freshwater N
266	source term (i.e. river inputs, land runoff); see below for more details. This
267	source is input into Box 2 (Winter Harbour; Fig 5).
268	• Mussel grazing, I_m , is partitioned amongst the boxes to correspond with present
269	conditions. Winter Harbour has no grazing (primarily a mussel spat collection
270	site) and I_m values for the other two boxes have been determined from the
271	nitrogen budget of the cultured mussel population (see below).
272	
273	Annual Nitrogen Budget. The average annual nitrogen inventories in Tracadie Bay
274	reservoirs were calculated as follows. N_{p} was calculated using the seasonal cycle of P
275	concentrations estimated from the objective analysis and the water volumes of the bay
276	(same procedure as described above for the boxes used in the lower trophic level model).
277	Nitrogen in the farmed mussel biomass was estimated based on a total harvested biomass
278	of approximately 1900 t wet weight (shell included). This number does not account for
279	mussel mortality, drop-off, or discarding of damaged or undersize mussels during
280	harvest. Estimates based on industry lease reporting place the biomass of mussels in the
281	bay at approximately 4500 t (Cranford et al. 2006). The nitrogen in mussel tissue
282	(excluding shell) was estimated using a typical fraction of wet meat to total weight of
283	40%, a water content of the meat of 85.5% (PJ Cranford unpublished data for Tracadie
284	Bay) and average nitrogen content of 7.79% (Smaal and Vonck 1997).
285	

286 The amount of phytoplankton and detritus nitrogen consumed by mussels depends on 287 the rate at which mussels filter water (i.e. their clearance rate) and the nitrogen content of 288 the suspended particulate matter in the water column. Clearance rate depends primarily 289 on the size of the mussels. We used a linear growth model to estimate the average 290 monthly size of Tracadie Bay mussels over a 24 month grow-out period to 0.7 g dry 291 weight at harvest. Meat weight trajectories in Tracadie Bay vary each year (Waite et al. 292 2005) and the linear function represents average conditions. The clearance rate (C) for 293 each mussel was calculated for each month using the allometric equation of Smaal et al. 294 (1997), which is based on similar natural dietary conditions as Tracadie Bay. Monthly 295 ingestion rates were calculated by multiplying C by the estimated number of mussels in the harvest (161 x 10⁶) and the average SPM-N concentration (2.5 mg SPM L^{-1} x 0.04 = 296 0.10 mg N L⁻¹). Nitrogen ingestion was summed over the 24 month period to estimate 297 298 annual ingestion. Note that summing monthly estimates from one cohort over a two year 299 period is equivalent to the actual situation where two cohorts (Age 1 and Age 2) are 300 present in the Bay each year. To estimate how much of the nitrogen in ingested food is 301 derived from phytoplankton and detritus, we assumed f_{POM} values of 0.8 and 0.2, respectively, for each food resource. Using the mean SPM value of 2.5 mg L^{-1} , and an 302 303 average annual seston f_{POM} value of 0.30 (SD = 0.14; n = 80; from the 2003 Niskin and *in* 304 situ water sampling survey), we estimate that approximately 40% of ingested seston 305 organic matter originates from phytoplankton. A similar proportion may be expected for 306 nitrogen ingestion. At the relatively low SPM concentrations found in Tracadie Bay, the 307 limited production of pseudofaeces does not significantly affect ingestion or 308 biodeposition estimates (Smaal et al. 1997; Cranford and Hill 1999), and is not 309 considered here.

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The amount of nitrogen in the faeces produced by the mussels can be determined from the difference between the nitrogen ingested and the nitrogen absorbed by the mussels. The latter depends on the absorption efficiency (*AE*) of the ingested food, which is in turn dependent on the concentration of organic matter in the SPM (f_{POM}). We estimated AE based on an empirical relationship between *AE* and f_{POM} ;

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$$AE = 0.85(1 - e^{-5^*(f_{POM} - 0.2)}),$$

derived from data reported for *M. edulis* by Cranford and Hill (1999) and Figueiras et al.

318 (2002). The average annual f_{POM} value of 0.30 (above) resulted in an AE value of 0.33.

319 Absorption rate was then calculated as the product of AE and nitrogen ingestion rate and

320 faeces production rate was calculated as ingestion rate minus absorption rate.

321

The annual excretion of nitrogen by mussels was estimated by two approaches. First, data presented in Hawkins and Bayne (1985) showed that, on average, 34% of absorbed nitrogen (calculated above) is excreted by mussels. Second, the allometric equation from Smaal et al. (1997) was used to estimate the excretion of NH_4 -N by different size classes of mussels in the same way as described above for estimating N ingestion. Most of the excreted nitrogen is in the form of dissolved ammonia.

328

Freshwater inputs used in the nitrogen budget are based on monthly averaged freshwater flow data for Winter River during 1968 to 2004, obtained from the Environment Canada hydrometric database (www.wsc.ec.gc.ca) for the station near Suffolk (46°19'56" N; 36°3'53"W; 37.5 km² drainage area). Average flow rate measurements from this station were adjusted by watershed areas not gauged to estimate the total monthly freshwater outflow from Winter River and the total freshwater run-off from all drainage sub-basins to Tracadie Bay. Data on nutrient concentrations from two sampling stations (surface water at WR1 and from 1 m depth at W1), both located near
the mouth of Winter River (Fig. 2), supplemented by estimates of levels during the winter
months for similar environments, were used along with the flow data for estimating TIN
fluxes in freshwater flowing into Tracadie Bay.

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341 Exchanges of nitrogen between Tracadie Bay and the offshore were estimated from 342 the tidal volume of the Bay and the concentrations of materials of interest in inflowing 343 and outflowing waters. The seasonal cycles for P, TIN and D were estimated for the 344 northern part of Tracadie Bay (defined as Box 1 of the lower trophic level model; Fig. 5) 345 and for offshore waters to characterize the outflow and inflow, respectively. These data 346 were combined with an estimate of 1.17 tidal volumes per day to yield gross estimates of 347 nitrogen export and import for each parameter. The number of tidal volumes per day was based on assessing flushing times for the bay by fitting a harmonic that describes the 348 349 three major components of the mixed tide (the O₁ and K₁ diurnal and the M₂ semi-350 diurnal) to spring and neap tides. 351

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Estimates for the average nitrogen inventories in Tracadie Bay and the internal andexternal annual nitrogen fluxes are shown schematically in Figure 6A.

RESULTS

Nitrogen Budget

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Reservoirs. We estimated an annual P inventory in Tracadie Bay of 1.2 t of nitrogen.

360 The equivalent inventory of TIN is 13 t N, and for detritus is 3.0 t N. The inventory of

nitrogen in mussel tissues in the Bay was estimated at 20 t N, with 9 t N y⁻¹ removed 361 362 annually in the mussel harvest. Our confidence in the former value is relatively low, so 363 the N budget (and the lower trophic level model) only considers the influences of a mussel biomass equal to the 9 t N y^{-1} harvest (Fig. 6A), for which there was reliable data. 364 The mussels harvested each year are estimated to ingest 230 t N v^{-1} , with 92 t N v^{-1} 365 366 originating directly from phytoplankton consumption. To the extent that this calculation 367 does not include a large, but poorly quantified, standing stock of cultured or wild 368 mussels, this ingestion rate should be interpreted as a lower limit. Applying a known 369 relationship for the absorption of organic matter by *M. edulis* resulted in estimates for absorption and faeces production rates of 76 t N y⁻¹ and 154 t N y⁻¹, respectively. Our two 370 estimates for nitrogen excretion provided comparable results, with 26 t N y⁻¹ calculated 371 based on the typical proportion of absorbed N that is excreted, and 23 t N v^{-1} derived by 372 373 applying the allometric equation of Smaal et al. (1997).

374

375 Freshwater Inputs. Monthly freshwater flows from Winter River over the period between 1964 and 2004 averaged between 0.5 (August - September) and 3.0 m³ s⁻¹ 376 (April), with an annual mean of $1.2 \text{ m}^3 \text{ s}^{-1}$. Scaling these flows to the remaining Tracadie 377 Bay watershed gave an average annual freshwater input of 2.6 m³ s⁻¹ (C.V. = 0.21). 378 379 Combining monthly average N concentrations at the mouth of Winter River with monthly water flows to Tracadie Bay vielded an estimate of 88 t N v⁻¹ for the annual freshwater 380 381 input of TIN. This estimate assumes that the water samples collected from the surface 382 layer had zero salinity (salinity data are not available for these samples). Since TIN 383 concentrations in Winter River are much higher than those in Tracadie Bay or offshore in 384 the Gulf of St. Lawrence, this estimate is a lower limit with respect to the presence of 385 some saltwater in the samples. The sub-surface (1 m depth) samples collected at the

adjacent sampling site (W1) had an average salinity of 27.1 psu. The freshwater
concentrations corresponding to the W1 samples were estimated using a two-component
mixing model, the average salinity (28.9 psu) at 15 m at the offshore station (W12), and
the monthly average TIN levels at W12 determined from the objective analysis of
offshore data as described above. The corresponding TIN fluxes based on these data are
equivalent to an annual flux of 124 t N y⁻¹.

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393 These two data-based estimates for the TIN flux can be compared to one based on

394 land use. Frink (1991) reviewed export coefficients for nutrients from watersheds to

395 estuaries and derived a model that predicted N coefficients (±SE) for agricultural,

forested and urban land types of 7.6 \pm 2.2, 2.4 \pm 0.5 and 13.4 \pm 2.6 kg N ha⁻¹ y⁻¹,

397 respectively. Combining these numbers with the corresponding land-use areas yields an average flux of 69 t N y⁻¹ from the Tracadie Bay watershed with a predicted range from 398 52 to 86 t N y⁻¹. N export from wetland areas was assumed to be minimal since they act 399 400 as N sinks (i.e. denitrification, sedimentation and plant uptake). Assuming an N coefficient of 8 kg N ha⁻¹ v^{-1} for atmospheric deposition (Frink 1991), the beach areas 401 would contribute approximately an additional 8 kg N y⁻¹, giving a total predicted TIN 402 flux of 76 kg N y⁻¹. The three estimates of total TIN inputs to Tracadie Bay are 403 reasonably consistent. For discussion purposes, we use a value of 100 t N y⁻¹ in our N 404 budget, which is based on the extensive Winter River measurements (flow and nutrient 405 406 concentrations) and includes some correction for nutrient dilution in river mouth samples by seawater. TIN input from agriculture is estimated to be 50 t N v^{-1} based on the 407 408 measured drainage basin land use and the above export coefficient for agriculture land. 409 Particulate nitrogen fluxes from land run-off are probably much less than the TIN fluxes 410 and have been assumed negligible in our budget.

412	Offshore Exchanges. Combining the seasonal cycles of phytoplankton (P) in
413	outflowing (northern Tracadie Bay) and inflowing (offshore) water with the daily tidal
414	flushing volume yielded a gross export of 122 t N y^{-1} and a gross import at 100 t N y^{-1} ,
415	which results in a net export of 22 t N y^{-1} . For TIN, gross export was 836 t N y^{-1} and
416	gross import was 183 t N y ⁻¹ , giving a net export of 654 t N y ⁻¹ . Tracadie Bay is a net
417	exporter of TIN in all months except January and November. The high winter TIN levels
418	in Tracadie Bay contribute substantially to the large net export. For detritus (D), gross
419	export was estimated to be 275 t N y^{-1} and gross import was 274 t N y^{-1} , for a net export
420	of 1 t N y^{-1} .
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422	Lower Trophic Level Model
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 425 426 427 428 429 430 431 432 	are summarized in Table 2. Computation of the maximum light limited phytoplankton growth rate,p(t), was based on a photosynthesis-irradiance relationship with the maximum photosynthetic rate modulated by temperature. A carbon to chlorophyll <i>a</i> ratio of 50 was used to convert the P-I relationship to a growth rate (see Dowd 2005). Daily values for the far field concentrations of $P_{\infty}(t)$, $N_{\infty}(t)$ and $D_{\infty}(t)$, and for nitrogen inputs, $N_{in}(t)$, into Winter Harbour (box 2) were derived using the objective analysis results described above. Values used for the N pool were total inorganic nitrogen (TIN = nitrate + nitrite + ammonia). Detritus inputs, $D_{in}(t)$, due to freshwater inputs and internal sources

437 Note that stochastic resuspension implies that the system will not repeat exactly year to

438 year and so annual fluxes may not exactly balance.

439

440 Some of the calculations for the mussel portion of the nitrogen budget are also used to 441 set parameters in the lower trophic level model. The model requires estimates for the 442 total filtration rate of the mussel population and the fractions of ingested nitrogen that are 443 harvested or excreted. Clearance rate of the annually harvested mussel biomass (9 t N y ¹) was determined during the ingestion rate calculation (above). Summing the clearance 444 rate over all size classes yields a total filtration rate of 6.3 x 10^6 m³ d⁻¹. Although detailed 445 446 stocking information that would allow calculation of the numbers of mussels in each 447 model box is not available, using the leased areas known to support mussel grow-out (as 448 opposed to leases used for spat collection) as proxies allocates ~ 92 % of the mussels to 449 box 1, ~ 1 % to box 2, and ~ 7 % to box 3. Scaling the total filtration rate to the volume of each box using these mussel densities produces ingestion rate, I_m , values of 0.29 d⁻¹ 450 for box 1 and 0.043 d^{-1} for box 3. We have set I_m equal to 0 for box 2 because of the 451 452 relatively clearance capacity of mussel spat. From the nitrogen budget, the excreted 453 fraction m = 26/230 = 0.11. Since the mussel portion of the budget is not fully balanced. 454 we have treated the assimilated fraction, m, as a tuneable parameter, and chosen its value so that the assimilated nitrogen matches the annual harvest of 9 Tons N y^{-1} . The resulting 455 value of m is 0.048. (Note that the assimilation efficiency is not the same as the 456 absorption efficiency, but they are related by AE = m + m. 457 458

459 Model Applications. The LTLM box model was applied to Tracadie Bay under460 three specific scenarios:

461 1. Cumulative Effect scenario representing the current state of Tracadie Bay with both 462 the cultured mussel population (M) and land-based TIN inputs at present day levels. 463 2. Enrichment Effect scenario without cultured mussels, but with land-based N inputs 464 at present levels. This scenario will be used to assess the impact of the cultured 465 mussels on the nitrogen dynamics of the ecosystem. 466 3. Baseline scenario without cultured mussels, and without land-based inputs of N. 467 This scenario tests the effect of the mostly agricultural land-based nitrogen source 468 on nitrogen dynamics. 469 470 The model outputs are presented in Figures 7-10 with each pane in these figures 471 showing the model predictions for the above three scenarios. In addition, the plots for P, 472 N and D (Figs. 7-10) show the observed concentrations (daily interpolations from the 473 objective analysis) in each box and the offshore concentrations. Table 3 lists the amount 474 of nitrogen in each reservoir and the annual fluxes between reservoirs as predicted by the

475 model for each box.

476

477 First, we can compare the model predictions for the present day scenario (cumulative 478 effects of mussels and nutrient enrichment) with the observations for P, N and D in each 479 of the model boxes. Both the P concentration ranges and the general seasonal patterns of 480 the P distributions predicted by the model are consistent with observations. The model 481 predicts spring and fall blooms in all three boxes, with summer values falling to 1-2 M N l⁻¹ (Fig. 7). However, the timing of the blooms predicted by the model are offset from 482 483 the observations by up to one month: e.g. the predicted spring blooms in box 1 and 3 are 484 about one month later than the observed blooms, while the predicted spring bloom in box 2 is approximately one month earlier. In addition, the fall blooms predicted by the model tend to be more intense than those observed. In the model, the highest spring P values occur in box 2 (Winter Harbour) due to the high N levels and lack of mussel grazing pressure, but observations show the highest values are in box 3 (head of Tracadie Bay).

490 The general spatial and temporal patterns in TIN (Fig. 8) conform with observations 491 with highest values in spring falling to near zero concentrations in summer, and smaller 492 increases early in fall that decline before the return of high values in the winter. Their 493 spatial pattern is also consistent (highest in box 2, then box 3, then box 1). However, the 494 predicted magnitudes are much smaller than the observations in spring. As previously 495 mentioned, the high TIN observations are based on a small number of samples collected 496 through the ice and gaps in the sampling occur in the spring. We do not have data to 497 indicate exactly when the high wintertime concentrations decrease, and whether or not 498 they contribute to spring productivity. The levels predicted by the model are consistent 499 with a typical temperate seasonal cycle, modified by high inputs into box 2 (especially 500 during the spring freshet).

501

The modelled water column detrital pool (Fig. 9) shows a fairly constant mean level near 5 _M N, with episodic fluctuations due to resuspension events, which is similar to both the observations in the individual boxes and to the levels offshore. The model also correctly predicts the shape and magnitude of the increase in D that occurs in box 2 in the spring.

507

508 The benthos in the model may be thought of as an ecologically active pool (or layer) 509 of nitrogen in which processes operate that result in resuspension, remineralization and

510	burial of nitrogen. The model predicts highest values for the cumulative effects scenario,
511	with box 1 (greatest biomass of mussels) containing the vast majority of benthic nitrogen.
512	The scenarios without mussels exhibited similar patterns and magnitude. Although the
513	benthos is an ecologically significant reservoir, there are no measurements suitable for
514	comparison with the model predictions shown in Fig. 10.
515	
516	DISCUSSION
517	
518	Lower Trophic Level Model Scenarios
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520	A comparison of estimated annual average phytoplankton levels in the different
521	model boxes for the Cumulative (mussels) and Enrichment (no mussels) scenarios (Table
522	3) show that mussel culture in Tracadie Bay affects all aspects of the nitrogen cycle to
523	some degree. Mussel grazing reduces phytoplankton levels by 15, 9 and 13% in boxes 1
524	(mouth of Tracadie Bay), 2 (Winter Harbour) and 3 (head of Bay), respectively. Although
525	the majority of mussels are located in box 1, phytoplankton depletion occurs throughout
526	the system owing to water exchange. The relatively large effect near the head of the bay,
527	despite the presence of only 7% of the cultured mussel population, results from poor
528	exchange with the offshore. Conversely, the impacts near the mouth are relatively small,
529	given that this area contains 92% of the cultured mussels, showing the importance of
530	offshore exchange and the supply of phytoplankton from Winter Harbour in regulating
531	phytoplankton levels. The conclusion that P is more reduced at the head of the bay than
532	in the mouth, is consistent with observations of reduced mussel growth near the head of
533	the bay (Waite et al. 2005), as well as theoretical studies considering the competing role
534	of P growth, M grazing, and the differential exchange processes (Dowd 2003).

536	Our predictions of reduced annual average P and D concentrations (13 and 14%
537	reductions, respectively) in the scenario with mussels (Table 3; Figs. 7 and 9) are
538	consistent with results from other studies of Tracadie Bay. Grant et al. (submitted) used a
539	more complex ecosystem model to investigate seston depletion and reported considerably
540	more severe effects of mussel culture on the overall P biomass than reported here. Those
541	predictions were validated using results from detailed surveys of the bay-wide
542	chlorophyll-a distribution. The density of second year mussels in the Grant et al.
543	(submitted) study (10 individuals m ⁻³) represents total mussel stocking levels and is
544	approximately double the value used in the current model application, which only
545	considers the effects of the harvested stock. The falling trend detected in the weight of
546	mussels harvested from Tracadie Bay during the 1990's, when annual stocking density
547	was steadily increasing (Figs. 4.3 and 4.4 in Cranford et al. 2006), implies a negative
548	feedback on meat yields caused by bivalve-induced food limitation. Over a 5-year period
549	when mussel biomass in the bay increased by more than 40%, the average mass yield of
550	mussel socks declined by 30%. Together, these observations and model results indicate
551	that the mussel production carrying capacity of Tracadie Bay has been exceeded.
552	

The pathways by which the nitrogen reaches the phytoplankton are dramatically altered in the presence of the farmed mussels. Mussel deposition (M _ B) sends 156 t N y⁻¹ to the benthos and the flux of nitrogen out of the sediments (B _ TIN + B _ D) is estimated to increase by 100 t N y⁻¹, enough to supply more than 70% of phytoplankton nitrogen requirements. In the presence of mussels, P _ D, D _ B, and D _ TIN fluxes are smaller in all three boxes (Table 3), presumably because mussel grazing consumes P and D that would otherwise be part of these fluxes. Asmus and Asmus (1991) raised the 560 possibility that mussels promote phytoplankton production by remineralizing detrital 561 material and by increasing rates of phytoplankton recycling during periods when N 562 demand is high and ambient concentrations are low. The model estimates that the mussels 563 ingest approximately twice as much detritus N than phytoplankton N (Fig. 6B; Table 3). 564 The recycling of detrital N through mussel excretion and biodeposition pathways will 565 promote phytoplankton growth during periods of N limitation and intensify bottom-up 566 controls on the phytoplankton. However, similar fluxes of TIN P in model runs with 567 and without mussels (Table 3) suggest no effect on annual phytoplankton production.

568

569 The impact of freshwater nitrogen inputs on annual average P levels in the different 570 boxes can be seen from a comparison of the Baseline (no mussels or inputs from land) 571 and Enrichment (no mussels) model runs (Table 3). As expected, freshwater inputs 572 increased TIN and P in all three boxes, with the greatest increases in Winter Harbour 573 (box 2), where the freshwater inputs occur. Removal of freshwater inputs resulted in P 574 reductions in Winter Harbour by as much as 62% during the spring bloom (Fig. 7), and 575 levels are reduced by as much as 50% in the remainder of the bay. Although these large P 576 reductions occur only in the spring (probably because the fall bloom is fuelled by 577 nutrients from offshore), the changes are large enough to have a sizeable impact on total annual phytoplankton growth (TIN P), which changes from 125 t N y^{-1} in the nutrient 578 enriched scenario to 72 t N v⁻¹ in the Baseline scenario (Table 3). Therefore, a substantial 579 580 fraction of the P present in Tracadie Bay is fuelled by land-derived nitrogen. P levels 581 outside Winter Harbour remained virtually the same as the current condition (when both 582 freshwater inputs and mussels are removed (compare the Cumulative and Baseline 583 scenarios in Table 3). These comparisons indicate; (1) the effect of freshwater nitrogen 584 on P levels is substantially greater than changes due to mussel grazing, and (2) cultured

586 food. The latter agrees with ecosystem model predictions of the large effect of watershed

587 nitrogen inputs on oyster production levels in the Thau lagoon (Chapelle et al. 2000).

588

585

589 Comparisons of box model results for scenarios with and without mussels suggest 590 that the presence of mussels increases retention of nitrogen from freshwater and offshore 591 sources within the bay. The mussels change the TIN export and M B and B TIN fluxes by 101, 156, and 91 t N y⁻¹ (increased by 2.3, 14 and 12 times, respectively). 592 593 Smaller changes (2.5 - 15%) also occur in the TIN P and D TIN fluxes when mussels 594 are present. Inspection of import / export terms in Table 3 shows that the TIN export from Tracadie Bay is much larger in the presence of mussels (176 t N y⁻¹) than in their 595 absence (75 t N y⁻¹), but this change is more than offset by the corresponding reduction in 596 P exports (51 t N v^{-1}) and the increase in D imports (78 t N v^{-1}). The combined effect of 597 598 all these changes is to produce slightly higher pelagic TIN levels and a much larger 599 benthic nitrogen pool when mussels are present than when they are not (Table 3; Figs. 8 600 and 10).

601

602 Dramatic changes in the relative role of the benthos in nitrogen cycling are apparent 603 in the presence of mussels. Mussel biodeposition is 3.5 times greater than the natural 604 sedimentation (D B) when mussels are not present (Table 3) and nitrogen burial increases by 72 t N y^{-1} , which is 11.8 times more than if no mussels were present. 605 Resuspension (B D) and remineralization (B TIN) increase by factors of 2.2 and 11.8, 606 607 respectively. As expected, the bulk of the benthic flux in the model run with mussels 608 occurs in box 1, where the majority of mussel grow-out takes place. The high B level in 609 this region represents a potential for severe eutrophication effects on benthic

610 communities. In contrast, the Enrichment model run indicated that freshwater inputs 611 have little impact on the nitrogen stored in the benthos (Table 3). It should be noted that 612 the model predictions of the fate of nitrogen after it reaches the benthos are only as good 613 as the model parameters controlling resuspension, remineralization and burial. Although 614 there are no field data to validate these specific estimates, model formulations of the 615 benthic component are based on robust equations of the important geochemical processes 616 involved (Dowd 2005). Model predictions are also parallel the results of a 2003 benthic 617 geochemical survey of Tracadie Bay that showed that hypoxic and anoxic sediment conditions, indicative of extensive organic enrichment, were only found within mussel 618 619 lease boundaries and that the majority (77%) of sampling sites with free sulphide 620 concentrations exceeding 1500 µM (13 sites) were located within box 1 (Cranford et al., 621 2006). Benthic macroinvertebrate communities throughout Tracadie Bay are described as having low diversity and a very low number of species (Miron et al. 2005). 622

623

624 Discrepancies between model predictions and observations (Figs. 7-10) could be due 625 to model errors in the forcing and far field conditions. They might also be due to errors of 626 representativeness in the point observations, e.g. a high productivity zone at the head of 627 Winter Harbour may have been under-sampled and so affect the seasonal cycles 628 constructed by objective analysis. Also critical to this comparison is that the model 629 scenarios are based on the influence of a mussel population equal to the amount harvested 630 each year. Although estimates are not well constrained, the total mussel biomass in the 631 bay appears to be double the annual harvest even without consideration of wild mussel 632 beds and ovster culture. The additional influences on nitrogen dynamics of large 633 populations of other herbivores residing on mussel culture structures would also need to 634 be modelled for a direct comparison between model output and observations to be valid.

636 aquaculture-induced changes to phytoplankton community composition.

637 Picophytoplankton cells, which are not captured by bivalves due to their small size, 638 appear to contribute substantially to phytoplankton biomass in extensively cultured 639 aquaculture embayments (Courties et al. 1994; Prins et al. 1998; Bec et al. 2005), 640 including Tracadie Bay (WKW Li personal communication; Cranford et al. 2006). The 641 philosophy of the parsimonious LTLM model has been to offer simplicity, but not 642 triviality, to quantitative descriptions of aquaculture systems by including only dominant 643 processes (Dowd, 2005). Although some realism is sacrificed for generality, the model 644 applications are based on robust parameterizations and approximations, well defined 645 boundary forcing, and data-driven estimation of mixing coefficients. We feel that the 646 resulting descriptions of observed parameters are therefore sufficient for the systematic testing of hypotheses concerning the effects of nutrient enrichment and mussel culture on 647 648 nitrogen dynamics (Dowd, 2005). 649 650 Nitrogen Budget

652 Biomass and fluxes involving mussels in the budget and the model cannot be 653 considered truly independent (calculated from similar underlying information) and will 654 not be compared. However, it is possible to directly compare some budget calculations 655 (Fig. 6A) with summed or averaged model estimates for the whole year for the combined 656 mussel and nutrient enrichment (Cumulative) scenario (Fig. 6B). Nitrogen inventories in 657 the different reservoirs from the budget and the model are similar except for TIN, for 658 which the model estimates are much lower (3.4 compared to 13 t N). A little less P (2.4 versus 22 t N y⁻¹) and a lot less TIN (176 versus 654 t N y⁻¹) are exported in the model 659

651

than in the budget, and a significant amount of D is imported in the model (106 t N y⁻¹), compared to D being in approximate balance in the budget (Fig. 6). The TIN levels and fluxes in the budget may have been biased by the high values obtained for the few available winter measurements. The budget has external nitrogen sources and sinks out of balance by 568 t N y⁻¹. In contrast, sources and sinks are nearly in balance for all three model scenarios (Table 3).

666

The nitrogen budget presented here was based on relatively simple concepts applied to some basic characteristics of Tracadie Bay and measurements or estimates of nitrogen levels in a few reservoirs, freshwater inputs, and relatively simple attempts to characterize the marine exchanges and fluxes associated with mussel feeding and excretion. Despite the simplifications, it was possible to derive the following noteworthy inferences on mussel/ecosystem interactions from the budget and associated calculations.

674 A comparison of the amount of nitrogen consumed by mussels with the inventories 675 of nitrogen in their food (P and D) and in the mussels themselves, shows that mussels 676 exert a dominant role in the flow of nitrogen through the Tracadie Bay ecosystem (Fig. 677 6A). The mussels ingest approximately 50 times the average standing stock of the total 678 nitrogen found in phytoplankton and detritus, which is equivalent to completely 679 processing the available food supply once a week. The mussels turn over plankton 680 nitrogen at an even higher rate (~5 days). Given that the mussel biomass in Tracadie Bay 681 is roughly double the amount used for this budget, the phytoplankton production 682 timescale would have to be on the order of a few days to support this level of aquaculture. 683 Dowd (2003) estimated a production timescale of 2-5 days for phytoplankton in 684 temperate coastal waters typical of Tracadie Bay during summer. Although primary

685 production data for this bay indicate remarkably high turnover times of 0.2-2.3 days (WG 686 Harrison, personal communication), the intensity of culture, in combination with the 687 other herbivores (including zooplankton, wild mussel reefs, some ovster culture and the 688 extensive fouling community on the mussel lines), is likely consuming available food at a 689 faster rate than can be replenished by internal production. Seston transported into the Bay 690 from offshore supplements the internal production. However, water residence time in 691 Tracadie Bay (3.4 days; Grant at al. 2005) is longer than the two days estimated for 692 mussel clearance of the tidal prism (Dowd, 2003). A high potential for bay-wide food 693 depletion is therefore indicated as was also concluded from the model results. 694 695 The budget cannot test hypotheses on the effects of mussels or freshwater inputs on 696 many important nitrogen reservoirs or internal fluxes, such as the biomass and 697 productivity of phytoplankton, ambient TIN levels and benthic storage of nitrogen. 698 Stated in other terms, the budget is not capable of testing the responses of Tracadie Bay 699 to forcing due to mussels, freshwater inputs or to different scenarios in general. Models 700 must be used to examine such internal processes and to test different scenarios. In 701 addition, the model, unlike the budget, provided spatial information resolved to the 702 geographic scale of the model boxes.

703

Although the budget has more limited application for testing hypotheses compared with the model, comparisons of nitrogen fluxes associated with the mussels with other fluxes in the budget (Fig. 6A) provide insights into potential pathways of aquaculture effects and have practical application. For example, a prevalent theory that can be addressed by the budget is that introduced bivalves modulate coastal eutrophication by clarifying the water and removing excess nitrogen in the harvest. The capacity for water 710 clarification has already been confirmed (see above). The ability of the bivalve harvest to 711 remove anthropogenic nitrogen inputs from land-use was examined by comparing freshwater inputs with the exported biomass. Mussel harvesting removes 9 t N v⁻¹ from 712 713 Tracadie Bay, which is equivalent to 9% of the total freshwater nitrogen input and 18% 714 of nitrogen input estimates from agricultural run-off. Given that phytoplankton accounted for 40% of the total ingested nitrogen (Fig. 6A), only 3.6 t N y⁻¹ of the mussel harvest 715 716 could result directly from phytoplankton uptake of agricultural nitrogen, with an 717 additional small amount removed via the P D M pathway (Fig. 6A). Therefore, only a 718 small fraction of the agricultural nitrogen run-off (<10%) appears to be removed by the 719 intensive mussel culture operations in this bay. Although the harvest does represent the 720 loss of a considerable amount of anthropogenic nitrogen from the coastal zone, intensive 721 bivalve aquaculture in Tracadie Bay does not appear to be an effective tool for coastal 722 managers to control nutrient emissions from land-use.

723

724 Mussel excretion and biodeposition represents significant fluxes in the nitrogen 725 budget (Fig. 6A) and must be considered along with grazing in any assessment of the 726 overall effects of aquaculture, including potential interactions with the effects of nitrogen 727 enrichment. Annual excretion by the harvested biomass returns an amount of nitrogen to 728 the water column equivalent to approximately 50% of the agricultural runoff, while 729 biodeposition was estimated to be three times greater than agricultural inputs (Fig. 6A). 730 Both fluxes from mussels were similar in magnitude to estimates from the lower trophic 731 level model (Fig. 6) and similar conclusion can be drawn on their potential ecologically 732 significance. Excretion by suspended mussels recycles nitrogen directly into nutrient 733 depleted waters. TIN levels in Tracadie Bay in late summer and fall, although low, are 734 dominated by ammonia, and silicate is generally the limiting nutrient for diatom growth

739 Our estimate of the flux of nitrogen to the sediments due to the harvested mussel biomass (154 t N v^{-1}) is equivalent to 105 mg N $m^{-2} d^{-1}$ if the deposition was uniformly 740 distributed over the $\sim 4 \text{ km}^2$ area of mussel grow-out leases and is approximately 200 741 mg N $m^{-2} d^{-1}$ when scaled up to the total mussel biomass in the bay (approximately twice 742 743 the harvested biomass). Grant et al. (2005) measured sedimentation rates in Tracadie Bay 744 and observed that rates under mussel lines were approximately double those at reference sites. An average biodeposition flux of 405 mg N m⁻² d^{-1} was estimated from Grant et al. 745 746 (2005) by subtracting natural sedimentation at the reference sites from total vertical flux 747 at the mussel sites and assuming a N content of 1.5% of dry faeces weight (Kautsky and 748 Evans 1987). This flux is higher than in our budget, as would be expected considering 749 that the sediment traps were placed close to the mussel lines, while our estimate also 750 includes areas between lines spaced ~5 m apart.

751

752 Reliability of Budget Estimates. A discussion of the nitrogen budget must consider 753 the confidence with which the budget estimates are determined. Three independent 754 means (two data-based and one based on nutrient export coefficients) were available for 755 estimating the freshwater inputs of TIN and they agreed to within \pm 26%, which makes 756 the freshwater input well known by the standards of these kinds of calculations. The 757 agricultural component of the freshwater input is based on applying published nutrient 758 export coefficients for nitrogen loading and comprehensive databases on land use and 759 river flow. Although nutrient loading will vary with the type of agriculture (Frink 1991), applying the known range of coefficients does not significantly affect our conclusions.

761

The nitrogen exchanges due to the mussels are expected to be reasonably constrained 762 763 by the basic physiology of the mussels. Although these fluxes are based on laboratory 764 measurements of individual mussels that were scaled up to an entire population in a real 765 environment, this is a common practice and the allometric relationships employed for 766 clearance rate provide results comparable with field measurements on mussel cohorts 767 (Cranford and Hill 1999). One test of the mussel component of the budget is to compare the total inputs and outputs of nitrogen to the mussels. The mussels consumed 230 t N v^{-1} . 768 and 189 t N v^{-1} of that consumption is accounted for in mussel production, urine and 769 770 faeces (Fig. 6A). Overall, nitrogen processed through mussels is balanced to within 771 $\sim 18\%$, which is within the bounds expected for this kind of a budget.

772

773 The offshore exchange estimates in the nitrogen budget must be considered first 774 order. Tidal prism calculations usually overestimate tidal transport because the 775 underlying assumptions of complete mixing within the Bay and infinite dilution outside 776 the Bay are approximations only. These calculations only consider exchanges due to the 777 tide and do not include exchanges due to other processes such as wind and storm events. 778 Applying the exchange coefficient for Tracadie Bay and the offshore, derived from a heat budget calculation ($K_{\infty} = 1.3 \text{ d}^{-1}$; Dowd 2005), resulted in net export estimates for P. TIN 779 and D of 28, 850 and 1.6 t N y⁻¹, respectively. These values are somewhat higher than 780 781 those predicted from the tidal prism (Fig. 6A), but they are within the same range. The budget estimates a large net export of all nitrogen forms from Tracadie Bay of 568 t N v⁻¹ 782 (= outputs – inputs). A large export value was predicted independent of whether the 783 784 tidal prism or heat budget exchange calculation was used, and is most likely due to the

785	high winter TIN concentrations that heavily influenced the TIN inventory. Despite a
786	potential for bias, the TIN inventory was only 13 t N and the calculations of the nitrogen
787	held in P and D are not subject to this uncertainty. Given these evaluations of confidence
788	in the different budget estimates, conclusions derived from the freshwater inputs and the
789	mussel processing of nitrogen are based on a more solid foundation than those derived
790	from the marine exchanges of nitrogen. The focus of the above discussion was therefore
791	on the former two aspects of Tracadie Bay nitrogen dynamics.
792	
793	CONCLUSIONS
794	
795	The following general conclusions about the cumulative influence of nutrient
796	enrichment and mussel aquaculture in Tracadie Bay were derived from the nitrogen
797	budget and lower trophic level model:
798	• mussels play a dominant role in nitrogen cycling in Tracadie Bay and influence all
799	aspects of the nitrogen cycle,
800	• a substantial fraction of the phytoplankton production in this inlet is fuelled by land-
801	derived nitrogen,
802	• the mussels depend on nitrogen in agricultural discharges to produce phytoplankton
803	biomass, as well as on phytoplankton and detritus (a major part of the food supply)
804	imported from offshore. That is, the internal production of the bay is insufficient to
805	support the harvested biomass of mussels,
806	• mussels are consuming available food at a faster rate than can be replenished by
807	internal and external processes. The budget and box model calculations indicate that
808	the productive capacity of Tracadie Bay for mussel aquaculture has been reached,

809	•	food may be less available to mussels at the head of the Bay than at the mouth,
810		despite the lower density of grow-out sites in the former location,
811	•	the amount of nitrogen removed in the mussel harvest is small (<10 %) compared to
812		the nitrogen in agricultural inputs,
813	•	mussels direct ~ 20 times more nitrogen to the water column and sediments in their
814		urine and biodeposits than is removed in the harvest and these processes increase
815		coastal nitrogen retention and dramatically alter pathways by which nitrogen reaches
816		the phytoplankton,
817	•	mussel biodenosition results in a very large flux of nitrogen to the benthos, with

mussel biodeposition results in a very large flux of nitrogen to the benthos, with
potentially serious eutrophication effects.

819

820 At the modeled level of mussel production, the pelagic ecosystem components are 821 much less sensitive to the effects of mussels than the benthic components, due to the 822 ability of external exchanges and feedbacks within the pelagic system to mitigate much 823 of the effect. However, the rapid depletion of external food supplies by mussels places 824 increased pressure on accelerated benthic remineralization processes and benthic/pelagic 825 fluxes to maintain the phytoplankton reservoir. Bivalve aquaculture utilizes nutrients 826 already present in the system, including those originating from land-use, and therefore 827 does not cause coastal nutrient enrichment. However, it does determine where 828 eutrophication from excess nutrient runoff is expressed by accelerating organic matter 829 transport to the seabed and thereby facilitating the retention of nutrients in the coastal 830 zone. Excessive benthic organic loading from biodeposition can lead to sediment anoxia. 831 Anoxic conditions inhibit nitrification and denitrification (Newell 2004) and 832 dissimilatory nitrate reduction to ammonium (Nizzoli et al. 2006) and would contribute to 833 a greater impact on N dynamics than estimated by the present model.

835	The nitrogen budget is a relatively simple representation of the marine exchanges and
836	fluxes associated with mussel aquaculture and terrestrial inputs in Tracadie Bay
837	compared with the dynamic ecosystem model. However, results from both approaches
838	are consistent and the lower trophic level model results do not contradict any important
839	conclusions from the budget. However, the finer resolution of the box model and the
840	ability to test different scenarios and predictions leads to additional conclusions. The
841	number of fundamental ecosystem processes influenced by mussels and the complexity
842	of their interactions makes it difficult to predict the impacts of mussels on many
843	ecosystem properties without resorting to a model. The detailed insight provided by a
844	dynamic model on ecosystem function can provide important information on issues like
845	suitable distributions of leases in an inlet that are beyond the scope of a budget.
846	
847	ACKNOWLEDGEMENTS
847 848	ACKNOWLEDGEMENTS
	ACKNOWLEDGEMENTS We gratefully recognize the contributions of many individuals involved in the
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848 849	We gratefully recognize the contributions of many individuals involved in the
848 849 850	We gratefully recognize the contributions of many individuals involved in the Tracadie Bay ecosystem study. S.L. Armsworthy and V. Burdette-Coutts provided field
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848 849 850 851 852	We gratefully recognize the contributions of many individuals involved in the Tracadie Bay ecosystem study. S.L. Armsworthy and V. Burdette-Coutts provided field and laboratory technical support for water sample collection and analysed the SPM and chlorophyll samples. D. Keen conducted the weekly sampling program in Winter River.
 848 849 850 851 852 853 	We gratefully recognize the contributions of many individuals involved in the Tracadie Bay ecosystem study. S.L. Armsworthy and V. Burdette-Coutts provided field and laboratory technical support for water sample collection and analysed the SPM and chlorophyll samples. D. Keen conducted the weekly sampling program in Winter River. C. Anstey analyzed the nutrient samples and G. Bugden provided some hydrographic
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Figure Legends
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Site maps showing Prince Edward Island, Canada, Tracadie Bay (including 860 Fig. 1. Winter Harbour), and the distribution of suspended mussel culture leases. 861 862 Fig. 2. Map of the Tracadie Bay watershed showing the locations of the five (A-E) land 863 drainage sub-basins, including the Winter River sub-basin (highlighted region 864 B). Pie charts presented for each sub-basin show the total area and percent landuse by categories as identified in the inset. Water sampling stations in Winter 865 River (squares) and inside and outside (Station W12) Tracadie Bay are shown. 866 867 Fig. 3. Seasonal cycle of nitrate in waters offshore from Tracadie Bay. Vertical lines 868 show the errors (± 1) associated with each monthly prediction. The histogram shows the monthly distribution of data points available for predicting the 869 870 seasonal cycle. Seasonal chlorophyll cycles predicted for the three boxes of the lower trophic 871 Fig. 4. 872 level model (Fig. 4), and the distribution of data used to make these estimates. 873 Map of Tracadie Bay showing the three boxes and boundaries used in the lower Fig. 5. 874 trophic level model. The intertidal and 2 m depth contours are also shown. 875 Fig. 6. Nitrogen reservoir inventories and flux pathway in Tracadie Bay calculated from (A) the nitrogen budget and (B) the "cumulative effects" scenario (present 876 877 conditions) of the lower trophic level model (LTLM). Both approaches are for 878 a mussel population equal to the annual mussel harvest. P = phytoplankton, TIN 879 = inorganic nitrogen, D = detritus, B = benthos, M = mussels. Solid arrows 880 represent internal fluxes and dashed arrows are external inputs/outputs. 881 Inventories in reservoirs are annual averages (t N) and all other quantities are annual fluxes (t N y⁻¹). For external fluxes, positive numbers represent gains to 882 883 Tracadie Bay and negative numbers are losses.

884	Fig. 7.	Lower trophic level model predictions for phytoplankton concentrations (P),
885		expressed as nitrogen equivalents. The model was run for three scenarios:
886		1) Cumulative = present day levels of mussels and freshwater nitrogen inputs,
887		2) Enrichment = no mussels, but freshwater nutrient input to the Bay, and
888		3) Baseline = no mussels present and no freshwater nitrogen input. The top of
889		the stippled area is the daily interpolated data for the offshore P concentration.
890		The top of the shaded area is the daily interpolated data for the observations in
891		each box.
892	Fig. 8.	Lower trophic level model predictions for nitrogen concentrations (TIN). The
893		model was run for the three scenarios described in Fig. 7. The top of the
894		stippled area is the daily interpolated data for the offshore TIN concentration.
895		The top of the shaded area is the daily interpolated data for the observations in
896		each box.
897	Fig. 9.	Lower trophic level model predictions for detritus concentrations (D), expressed
898		as nitrogen equivalents. The model was run for the three scenarios described in
899		Fig. 7. The top of the stippled area is the daily interpolated data for the offshore
900		D concentration. The top of the shaded area is the daily interpolated data for the
901		observations in each box.
902	Fig. 10.	Lower trophic level model predictions for nitrogen levels in the benthos (B).
903		The model was run for the three scenarios described in Fig. 7.
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Symbol	Description	N Budget	Lower Trophic Level Model		
Reservoirs					
Р	Phytoplankton	У	у		
TIN	Inorganic nitrogen	y	y		
D	Detritus	y	y		
В	Benthos	n	y		
М	Cultured mussels	У	Specified		
Internal Fluxe	S				
TIN _ P	Photosynthesis	n	у		
P_D	Mortality	n	y		
D_TIN	Remineralization (water column)	n	y		
B_TIN	Remineralization (benthos)	n	у		
D_B	Sedimentation	n	У		
B_D	Resuspension	n	у		
P_M	Ingestion	У	У		
D_M	Ingestion	У	У		
M_B	Biodeposition	У	У		
M_TIN	Excretion	У	У		
External Flux	es				
M Harvest		У	У		
River	Winter River discharge	TIN only*	TIN only*		
Offshore	Marine exchange	P, TIN and D	P, TIN and D		
Burial		n	У		
Atmospheric	Denitrification	n	n		

Table 1. Elements of the nitrogen cycle in Tracadie Bay. Quantities calculated or predicted by each model are indicated by 'y' and those not considered by 'n'. P = phytoplankton, TIN = inorganic nitrogen, D = detritus, B = benthos, and M = mussels.

*P and D river discharge assumed negligible

Table 2. Definition of quantities in the lower trophic level model. Groupings are according to variable type. For each quantity the following information is given: units, its numerical value (or its source), and a brief definition. Explicit functional dependence on time (t) or temperature (Temp) is indicated. P = phytoplankton, TIN = inorganic nitrogen, D = detritus, B = benthos, and M = mussels. Other symbols are defined in Dowd (2005).

Quantity	Units	Value	Definition
(i) State Variables			
Р	mmol N m ⁻³	See text	phytoplankton
Ν	mmol N m ⁻³	See text	nutrients
D	mmol N m ⁻³	See text	water column detritus
В	mmol N m ⁻²	See text	benthic detritus
(ii) Parameters			
K	d^{-1}	Dowd (2005)	exchange/flushing coefficient
k _n	mmol N m ⁻³	2.5	half-saturation for N uptake by P
_p(t)	0.2 - 1	eq. (8), Dowd (2005)	P growth rate
p	mmol N m $^{-3}$ d $^{-1}$	0.05	grazing loss of P
_d(Temp)	d^{-1}	0.02 - 0.1	remineralization rate of D to TIN
d	d^{-1}	0.05	sinking rate for D
_b(Temp)	d^{-1}	0.01	remineralization rate for B to TIN
r(t)	mmol N m $^{-3}$ d $^{-1}$	varies	resuspension flux
_	-	0.01	burial fraction
Īm	d^{-1}	see text	ingestion rate of bivalves
m	-	0.065	assimilated fraction for bivalves
m	-	0.11	excreted fraction for bivalves
	m	4	water depth
(iii) External Inpu	ts		
$P_{\infty}(t)$	mmol N m ⁻³	see Fig	far-field P
$N_{\infty}(t)$	mmol N m ⁻³	see Fig	far-field TIN
$D_{\infty}(t)$	mmol N m ⁻³	see Fig	far-field D
$N_{in}(t)$	mmol N m $^{-3}$ d $^{-1}$	varies	external TIN input
$D_{in}(t)$	mmol N m ^{-3} d ^{-1}	0	external D input

Table 3. Nitrogen inventories (annual averages, t N) and fluxes (t N y⁻¹) calculated from the lower trophic level model. Model scenarios are defined in the text. 'Exchange' terms for boxes 1-3 are the net exchanges with other boxes or the offshore (positive terms are a net gain to the box). The 'Total' column is the exchange between box 1 and the offshore, and represents the material lost or gained in all of Tracadie Bay by marine exchange. P = phytoplankton, TIN = inorganic nitrogen, D = detritus, B = benthos, and M = mussels.

-	Cum	ulate Effe	cts (Scena	rio 1)	Enrichment Effects (Scenario 2)				Baseline (Scenario 3)			
	Box 1	Box 2	Box 3	Total	Box 1	Box 2	Box 3	Total	Box 1	Box 2	Box 3	Total
Р	0.55	0.43	0.33	1.31	0.65	0.47	0.38	1.51	0.57	0.33	0.32	1.23
TIN	1.45	1.24	0.71	3.40	1.24	1.15	0.59	2.98	0.97	0.49	0.49	1.96
D	1.08	0.56	0.50	2.14	1.25	0.65	0.59	2.48	1.18	0.53	0.54	2.24
В	8.02	0.11	0.30	8.43	0.36	0.13	0.13	0.62	0.34	0.10	0.12	0.56
TIN _ P	47	48	28	122	48	49	28	125	32	20	20	72
P_D	18.4	25	13.0	56	26	29	17.3	72	19.4	12.8	12.2	44
D_TIN	40	20	18.2	78	46	23	21	90	44	19.2	20	82
B_TIN	92	1.9	4.4	99	4.2	2.3	2.0	8.4	3.9	1.78	1.79	7.5
D_B	20	10.2	9.1	39	23	11.8	10.8	45	21	9.6	9.8	41
B_D	13.6	2.0	3.7	19	4.4	2.3	2.1	8.8	4.2	1.88	1.90	8.0
P_M	58	0	5.1	63	0	0	0	0	0	0	0	0
D_M	114	0	8	122	0	0	0	0	0	0	0	0
M_B	145	0	11	156	0	0	0	0	0	0	0	0
M_TIN	19.0	0	1.4	20.4	0	0	0	0	0	0	0	0
M harvest	8.3	0	0.6	8.9	0	0	0	0	0	0	0	0
TIN River	0	101	0	101	0	101	0	101	0	0	0	0
P Exchange	30.1	-22.9	-9.5	-2.4	-22	-21	-10.3	-53	-13.0	-7.4	-7.5	-28
TIN Exchange	-105	-75.2	3.6	-176	-2.1	-77	4.5	-75	-14.9	-0.80	-1.67	-17.4
D Exchange	101	-2.8	7.4	106	25	-3.2	6.1	28	29	8.5	9.5	47
burial	74	1.53	3.5	79	3.3	1.81	1.60	6.7	3.1	1.43	1.44	6.0



Fig. 1.



* Water sampling stations: WR1 (left square) and W1 (right circle).

Fig. 2.



Fig. 3.



Fig. 4.



Fig. 5.







Fig. 7.



Fig. 8.







Fig. 10.